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#### **Product Datasheet**

# Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]

# <u>Applications</u> | <u>Antibody Details</u> | <u>Antigen Details</u> | <u>References</u>

#### Overview

Antigen Glypican 3 (GPC3)

Immunogen Purified recombinant fragment of Mouse Glypican 3 (GPC3), AA: 25-553.

Host/isotype Rabbit/IgG1

Clonality Recombinant monoclonal

Clone name IPI-mGPC3.19 RRID AB\_3101862

IPI ID TAB0010792-013-002

Specificity GPC3; Does not recognize other GPCs

Species reactivity Mouse and human

Amount 100  $\mu$ g Concentration 1 mg/mL

Purification Expressed in ExpiCHO cells and affinity purified using Protein A

Storage buffer PBS, pH 7.4 with 0.02% ProClin 300

Shipping Shipped on blue ice at +4C

Storage Store at +4C for up to 3 months. For long-term storage, aliquot and store at -20C.

Avoid multiple freeze/thaw cycles.

# **IPI Tested Applications**<sup>‡</sup>

Application	Tested concentration	Result	Reference
Cell Display	5 μg/mL	Positive	https://doi.org/10.57733/addgene.fndotl
Flow	5 μg/mL	Positive	https://doi.org/10.57733/addgene.16x4s7
IF	1 μg/mL	Positive	https://doi.org/10.57733/addgene.nlepjk
WB	1 μg/mL	Negative	Unpublished

<sup>&</sup>lt;sup>‡</sup> Not suitable for WB application.

#### Community Data\*

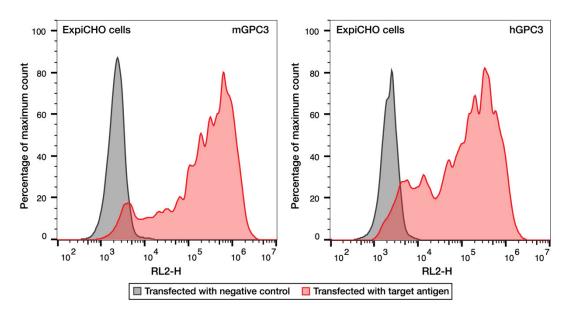
Application	Lab	Reference
IF	Adrian Salic, Ph.D., Harvard Medical School	https://doi.org/10.57733/addgene.somgvn

<sup>\*</sup> Supporting Data is generated by external partner labs, in the process of evaluating IPI antibody panels.

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# **Applications**

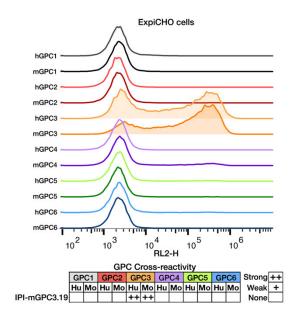
## **Cell Display**



Anti-Glypican 3 (GPC3) [IPI-mGPC3.19] recognizes mouse and human GPC3. Histogram from FACS analysis on ExpiCHO cells transfected with mouse or human glypican 3 (red), or WY0041 negative control (gray). Cells expressing mouse (left panel) or human (right panel) GPC3 were labeled with Anti-Glypican 3 (GPC3) [IPI-mGPC3.13] (Addgene #222515) and Alexa Fluor 647 F(ab')2 Fragment Goat Anti-Rabbit IgG Fc (Jackson ImmunoResearch, 111-606-046). Labeled cells were analyzed with an Intellicyt iQue Screener Plus flow cytometer. Histograms were generated and normalized to mode using FlowJo™ v10.10. doi: https://doi.org/10.57733/addgene.fndotl

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#### Flow cytometry

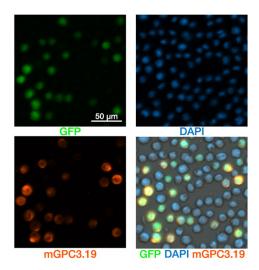


Histogram overlays from FACS analysis on ExpiCHO cells transfected with human and mouse glypicans (GPC1-GPC6). Cells expressing GPCs were labeled with Anti-Glypican 3 (GPC3) [IPI-mGPC3.19] (Addgene #222515) and Alexa Fluor 647 F(ab')2 Fragment Goat Anti-Rabbit IgG Fc (Jackson ImmunoResearch, 111-606-046). Labeled cells were analyzed with an Intellicyt iQue Screener Plus flow cytometer. Anti-Glypican 3 (GPC3) [IPI-mGPC3.19] is specific for human and mouse GPC3 (orange lines with fill) and does not recognize other GPCs. Histograms were normalized to mode and displayed as an overlay with half offset using FlowJo™ v10.10. Bottom table summarizes GPC cross-reactivity. Abbreviations: glypican (GPC), human (Hu), mouse (Mo), strong (++), weak (+), none ( ). doi: https://doi.org/10.57733/addgene.16x4s7



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#### **Immunofluorescence**



Immunofluorescence (IF) of ExpiCHO cells transfected with human GPC3. ExpiCHO cells were transiently transfected with GFP (transfection control) and human GPC3. 100,000 cells were fixed to a 96-well glass bottom plate using 4% PFA at 37C for 15 min. Cells were permeabilized with 0.1% Triton X-100 at RT for 10 min and then blocked with 5% BSA, 5% serum, and 0.01% Triton X-100 at room temperature for 30 min. Cells were incubated with 1 μg/mL Anti-Glypican 3 (GPC3) [IPI-mGPC3.19] (Addgene #222515). Alexa Fluor 647 F(ab')2 fragment goat anti-rabbit IgG, Fc fragment-specific (Jackson ImmunoResearch, 111-606-046) was used as the secondary antibody. Nuclear staining was performed using 0.5 mg/mL DAPI. Cells were imaged for GFP (green), DAPI (blue), and GPC3 (red) using an EVOS M7000 microscope at 10x magnification. doi: https://doi.org/10.57733/addgene.nlepjk

# Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]



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#### **Antibody Details**

#### Antibody design and production

Human variable domains for the heavy and light chain of the FAB fragment used in yeast display were grafted onto the constant CH1, CH2 and CH3 domains of rabbit IgG. The chimera human/rabbit IgG1 construct was recombinantly expressed in ExpiCHO cells, using pTipi2.1 as the expression vector. The antibody was purified by affinity chromatography using protein A (XYZ) and acid elution, followed by immediate buffer exchange using 1 x PBS buffer pH 7.4.

## **Sequence information**

Heavy chain and light chain amino acid sequences are available upon request after purchase. Contact us to request.

## **Antibody Characterization**

**LC-MS:** Intact mass analysis via LC-MS methods allows for confirmation antibody mass, and to identify any product-related variants such as glycosylation. Before conducting intact mass analysis via LC-MS, the antibody was reduced to its heavy chain (HC) and light chain (LC). This process allows for confirmation of the masses corresponding to the amino acid sequences of both chains.

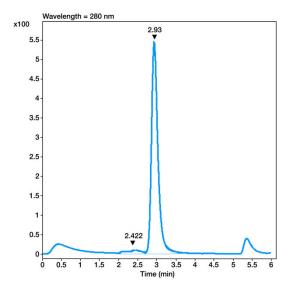
	HC MW (Da) Calculated	HC MW (Da) Observed	HC MW (Da) Delta	LC MW (Da) Calculated	LC MW (Da) Observed	LC MW (Da) Delta
IPI-mGPC3.19	49667.53	49672.70	5.17	22944.51	22944.31	-0.20

**Heavy Chain (HC) Mass Calculation:** The calculated molecular weight (MW) of the HC is derived by adding the mass of the unmodified HC amino acid sequence to the mass of the predominant N-glycan form (G0F), which is 1444.5 Da. This calculation assumes that the intrachain disulfide bonds remain intact. For HCs with an N-terminal glutamine (Q), the mass of Q is converted to pyroglutamic acid (PyroGlu), resulting in a deduction of 17.03 Da from the total mass. Additionally, for HCs with a C-terminal lysine (K), the mass of K (128.09 Da) is also subtracted.

**Light Chain (LC) Mass Calculation:** The calculated molecular weight (MW) of the LC is obtained from the mass of the unmodified LC amino acid sequence, assuming that the intrachain disulfide bonds are not reduced. For LCs with an N-terminal glutamine (Q), the mass of Q is converted to pyroglutamic acid (PyroGlu), leading to a deduction of 17.03 Da from the total mass. For LCs with a C-terminal lysine (K), the mass of K (128.09 Da) is subtracted as well.

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**Size Exclusion Chromatography (SEC):** SEC is a protein purification technique that separates molecules based on size.



High-performance liquid chromatography-size exclusion chromatography (HPLC-SEC) profile. 5  $\mu$ g of antibody (1 mg/mL) was analyzed. 1x PBS, pH 7.4 was used as the mobile phase. Protein standards ranging from 15-600 kDa were used for calibration (Sigma, 69385) and Lox1.9 antibody was run as an internal control. Resulting peaks were analyzed using Agilent OpenLab CDS software to determine total peak area and percentage purity.

	Purity	Result
IPI-mGPC3.19	≥95%	Pass

# Institute for Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]

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## **Antigen Details**

## Immunogen design:

cDNA of Mouse Glypican 3 (GPC3) with C-terminal His-, Avi-, and HA-tags was produced in transiently transfected Expi293F cells and purified from culture supernatant by Ni-NTA affinity purification followed by size-exclusion chromatography.

#### Immunogen sequences:

Mouse GPC3 (AA: 25-553)

QPPPPPDATCHQVRSFFQRLQPGLKWVPETPVPGSDLQVCLPKGPTCCSRKMEEKYQLTARLNMEQLL QSASMELKFLIIQNAAVFQEAFEIVVRHAKNYTNAMFKNNYPSLTPQAFEFVGEFFTDVSLYILGSDINVDD MVNELFDSLFPVIYTQMMNPGLPESVLDINECLRGARRDLKVFGSFPKLIMTQVSKSLQVTRIFLQALNLG **IEVINTTDHLKFSKDCGRMLTRMWYCSYCQGLMMVKPCGGYCNVVMQGCMAGVVEIDKYWREYILSLE** ELVNGMYRIYDMENVLLGLFSTIHDSIQYVQKNGGKLTTTIGKLCAHSQQRQYRSAYYPEDLFIDKKILKV AHVEHEETLSSRRRELIQKLKSFINFYSALPGYICSHSPVAENDTLCWNGQELVERYSQKAARNGMKNQF NLHELKMKGPEPVVSQIIDKLKHINQLLRTMSVPKGKVLDKSLDEEGLESGDCGDDEDECIGSSGDGMV KVKNQLRFLAELAYDLDVDDAPGNKQHGNQKDNEITTSHSGSGHHHHHHHHHHHGSGGLNDIFEAQKIE WHEGSGDYKDDDDK

#### Sequence information:

HUGO: MGI:104903 Uniprot: Q8CFZ4 Refseq: NM 016697.3

## Structural information:

Topology: Glycosylphosphatidylinositol (GPI) Anchored

PDB IDs: <u>7ZA1, 7ZA2, 7ZA3, 7ZAV</u>

Alphafold: AF-Q8CFZ4-F1

#### **Expression profiles:**

Human Protein Atlas ENSG00000147257-GPC3

# Protein Innovation

# Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]

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## References

- 1. Z. Anderson, H. Li, T. Riedel, H Zhu, R. Meijers, and D. Moshinsky. (2024). Flow Cytometry for Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]. Addgene. <a href="https://doi.org/10.57733/addgene.fndotl">https://doi.org/10.57733/addgene.fndotl</a>
- 2. Z. Anderson, H. Li, T. Riedel, H Zhu, R. Meijers, and D. Moshinsky. (2024). Flow Cytometry for Anti-Glypican 3 (GPC3) [IPI-mGPC3.19]. Addgene. <a href="https://doi.org/10.57733/addgene.16x4s7">https://doi.org/10.57733/addgene.16x4s7</a>
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- 4. A. Salic. (2024). Immunofluorescence for Anti-glypican 3 (GPC3) [IPI-mGPC3.19]. Addgene. <a href="https://doi.org/10.57733/addgene.somgvn">https://doi.org/10.57733/addgene.somgvn</a>

# How to cite this antibody:

Anti-Glypican 3 (GPC3) [IPI-mGPC3.19] - from Institute for Protein Innovation (IPI) (Addgene #222515; http://n2t.net/addgene:222515; RRID: AB\_3101862).

If you publish research with this product, please let us know so that we can cite your paper.